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LEAD-210 AND POLONIUM-210 IN WATER

(WITH VACUUM BOX SYSTEM)

1. SCOPE

- 1.1. This is a method for the separation and measurement of lead-210 and polonium-210 in water.
- 1.2. This method does not address all aspects of safety, quality control, calibration or instrument set-up. However, enough detail is given for a trained radiochemist to achieve accurate and precise results for the analysis of the analyte(s) from the appropriate matrix, when incorporating the appropriate agency or laboratory safety, quality and laboratory control standards.

2. SUMMARY OF METHOD

2.1. Lead and polonium are concentrated from up to 1.5L samples by iron hydroxide precipitation. Pb and Po are separated from the other elements with Eichrom Sr Resin. The purified lead fraction is prepared for analysis evaporation onto a stainless steel counting planchet or by liquid scintillation counting. The polonium fraction is prepared for alpha spectrometry by autodeposition onto a metal planchet or by bismuth phosphate precipitation onto an Eichrom Resolve[®] Filter.

3. SIGNIFICANCE OF USE

3.1. This is a rapid, reliable method for measurement of lead and polonium in water samples that is efficient and reduces interferences to analysis.

4. INTERFERENCE

- 4.1. Any beta emitter can cause an interference with the measurement of the ²¹⁰Bi daughter of ²¹⁰Pb.
- 4.2. Polonium auto deposition onto silver, copper or nickel planchet can be reduced by high concentrations of Fe, Te, Au, Pt and Hg.
- 4.3. This method effectively eliminates these possible interferences.

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5. APPARATUS

- Alpha Spectrometry system (For measurement of Po)
- Aluminum foil 0.003 inch thickness, if counting ²¹⁰Bi and ²¹⁰Pb via gas flow proportional counter.
- Beakers, glass
- Centrifuge
- Centrifuge tubes, plastic, 50mL
- Detection System for measurement of ²¹⁰Pb via ²¹⁰Bi daughter:
 - Option 1: Low background proportional counter
 - Stainless steel planchets, cupped flat bottom, compatible with sample holders for proportional counter
 - Option 2: Liquid Scintillation Counter (LSC)
 - Liquid scintillation cocktail
 - Liquid scintillation vials, 20mL
- Fume hood
- Hot plate
- Planchets, Silver, copper or nickel, flat, 1 inch diameter, polished on one side and coated on the back side with an acid resistant acrylic paint.
- Resolve[™] filter- 0.1 micron 25 mm polypropylene, Eichrom Part: RF-100-25PP01 and filter apparatus- Pall 25mm polysulfone filter apparatus with polycarbonate base, metal screen and 50 mL reservoir (Pall Part: 4203)

-or-

- Resolve[™] filter- 0.1 micron 25 mm polypropylene in disposable funnel, Eichrom Part: RF-DF-25-25PP01
- Stir rods, glass
- Vacuum box, Eichrom Part: AR-BOX-12 or AR-BOX-24
- Inner support tubes, Eichrom Part: AR-1000-TUBE-PE
- Vacuum pump, dry pump, 115V, 60 Hz, Fisher Part: 01-092-25 or equivalent
- Water bath, shaking or ultrasonic
- Yellow outer tips, Eichrom Part: AR-1000-OT

6. REAGENTS

Note: Analytical grade or ACS grade reagents are recommended. Evaluation of key reagents, for contribution to method background levels from naturally occurring radioactive materials is recommended.

Ammonium citrate, $(NH_4)_3C_6H_5O_8$
Ammonium Hydroxide (56% NH₄OH or 28% NH₃), conc. NH₄OH
Ammonium Hydrogren Phosphate, (NH ₄) ₂ HPO ₄
Bismuth Chloride, BiCl ₃
Sodium Hydroxide, NaOH
Ascorbic acid, C ₆ H ₈ O ₆
Deionized water, All reagents are prepared with deionized water
Ethanol denatured, C ₂ H ₅ OH
Ferric nitrate nonahydrate, Fe(NO ₃) ₃ .9H ₂ O
Hydrochloric acid (37%), concentrated HCI
Isopropyl alcohol C ₃ H ₇ OH
Lead nitrate, Pb(NO ₃) ₂
<i>Nitric acid (70%),</i> concentrated HNO ₃
Phenolphthalein pH Indicator
Po yield tracer, ²⁰⁸ Po or ²⁰⁹ Po
Sr [®] resin, 2mL prepacked cartridge, 50-100µm, Eichrom Part SR-R50-S
Sulfuric acid (98%), concentrated H ₂ SO ₄

- 6.1. Ammonium Citrate (0.05M) Add 11.3g of ammonium citrate and 700mL water to a 1L volumetric flask. Mix to dissolve. Dilute to 1L with water.
- 6.2. Ammonium Hydrogen Phosphate (3.2M) Dissolve 211g (NH₄)₂HPO₄ in 400mL water. Dilute to 500mL with water.
- 6.3. Ascorbic Acid (1M) Dissolve 17.6g of ascorbic acid in 90mL of water. Dilute to 100mL with water. **Prepare fresh weekly.**
- 6.4. *Hydrochloric Acid (0.5M)* Add 41mL concentrated HCI to 800mL water. Dilute to 1L with water.

- 6.5. *Hydrochloric Acid (2.0M)* Add 165 mL concentrated HCl to 700 mL water. Dilute to 1L with water.
- 6.6. *Iron carrier* (20 mg/mL) − Dissolve 14.5g of Fe(NO₃)₃·9H₂O in 70mL water. Dilute to 100mL with water.
- 6.7. Lead carrier (10 mg/mL) Dissolve 1.6 grams of Pb(NO₃)₂ in water and dilute to 100mL with water (Check batches for ²¹⁰Pb and ²¹⁰Po levels).
- 6.8. Bismuth carrier (1mg/mL) Dissolve 0.75g BiCl₃ and 42mL 37% HCl in 200mL of water. Dilute to 500mL with water.
- 6.9. *Nitric acid solution (8M)* Add 500 mL of concentrated HNO₃ to 400mL of water. Dilute to 1L with water.
- 6.10. *Nitric Acid (1M) A*dd 62.5 mL of concentrated HNO₃ to 800mL water. Dilute to 1L with water.
- 6.11. *Nitric Acid (0.1M)* Add 6.3 mL of concentrated HNO₃ to 800mL water. Dilute to 1L with water.
- 6.12. *Phenolphthalein solution* Dissolve 1g phenolphthalein in 100mL 95% isopropyl alcohol. Dilute with 100mL of water.
- 6.13. Sodium Hydroxide (1M) Dissolve 40g of NaOH in 800mL of water. Dilute to 1L with water.

7. PROCEDURE

- 7.1. Water Sample Preparation:
 - 7.1.1. If required, filter the sample through a 0.45 micron filter.
 - 7.1.2. Aliquot 500 to 1,500 mL of the sample (or enough to meet required detection limit) into an appropriate size beaker.
 - 7.1.3. Acidify the sample to pH 2, with concentrated HNO₃.
 - 7.1.4. Add 1 mL of stable Pb carrier or an appropriate amount of Po tracer.
 - 7.1.5. If samples larger than 1L are analyzed, evaporate the sample to approximately 1L.
 - 7.1.6. Iron hydroxide precipitation
 - 7.1.6.1. Add 1mL of iron carrier to each sample. Cover beaker with a watch glass and heat at near boiling for an hour.
 - 7.1.6.2. Remove watch glass. Add 1mL of phenolphthalein solution. While stirring, slowly add enough concentrated



NaOH to reach the phenolphthalein pink endpoint (pH 8-9). An iron hydroxide precipitate should form. Allow the beakers to heat for another 30 minutes.

Note: NaOH should be used to form the ferric hydroxide ppt. Adjusting the pH with NH₄OH can lead to incomplete precipitation of iron and lower Pb/Po recoveries.

- 7.1.6.3. Let the precipitate settle for 1-2 hours or centrifuge.
 Decant the supernate. Transfer the precipitate into a 50 mL plastic centrifuge tube with deionized water.
- 7.1.6.4. Centrifuge and discard the supernate.
- 7.1.6.5. Wash the precipitate with 10mL water. Centrifuge and discard the supernate.
- 7.1.6.6. Add 10mL 2M HCl to dissolve the precipitate.
- 7.1.6.7. Add 1mL of 1M ascorbic acid to each solution, swirling to mix. Wait for 2-3 minutes.

Note: Any yellow color should disappear which indicates the reduction of Fe(III) to Fe(II). If the color persists add ascorbic acid should drop-wise with mixing until the color disappears. Add 0.5mL excess ascorbic acid to maintain Fe reduction.

- 7.2. Pb and Po Separation using Sr Resin
 - 7.2.1. Place the inner tube rack (supplied with vacuum box system) into the vacuum box with the centrifuge tubes in the rack. Fit the lid to the vacuum system box. Alternatively, a vacuum box inner liner may be used.
 - 7.2.2. Place yellow outer tips into all 12 or 24 openings in the lid of the vacuum box. Fit a white inner support tube into each yellow tip.
 - 7.2.3. For each sample solution, fit a Sr Resin cartridge on to the inner support tube.
 - 7.2.4. Add syringe barrels (funnels/reservoirs) to the top end of each Sr Resin cartridge.
 - 7.2.5. Connect the vacuum pump to the box. Turn the vacuum pump on and ensure proper fitting of the lid.



Note: The unused openings on the vacuum box should be sealed. Vacuum manifold plugs can be used to plug unused white tips to achieve good seal during the separation. Alternatively, unused vacuum box holes can be sealed with scotch tape.

- 7.2.6. Add 10mL 2M HCl to each Sr Resin cartridge reservoir to condition the resin. Adjust vacuum to achieve a flow rate of 1-2mL/min. Allow solution to completely pass through each cartridge.
- 7.2.7. Load the sample onto the Sr resin. Allow solution to completely pass through each cartridge at 1-2mL/min.
- 7.2.8. Rinse each sample tube with 5mL 2M HCI. Add the rinse to each Sr Resin cartridge reservoir. Allow solution to completely pass through each cartridge at 1-2mL/min.
- 7.2.9. Place a clean reservoir above each Sr Resin cartridge.
- 7.2.10. Rinse each Sr Resin cartridge with 5mL 2M HCI. Record the time and date of this addition. This will be used to calculate the ingrowth of ²¹⁰Bi. Allow solution to drain. Allow solution to completely pass through each cartridge at 1-2mL/min. Disengage vacuum. Discard the eluent as waste.

Note: This will remove bismuth and yttrium if present.

- 7.2.11. Place a clean, labeled centrifuge tube under each Sr Resin cartridge to collect Polonium fractions.
- 7.2.12. Add 5mL 1.0M HNO₃ to each Sr Resin cartridge reservoir. Engage vacuum. Allow solution to completely pass through each cartridge at 1mL/min.
- 7.2.13. Add 15mL 0.1M HNO $_3$ to each Sr Resin cartridge reservoir. Allow solution to completely pass through each cartridge at 1mL/min.
- 7.2.14. Disengage vacuum. Remove tubes and set aside for Po autodeposition in Section 7.3.
- 7.2.15. Place a clean, labeled centrifuge tube under each Sr Resin cartridge to collect Pb fraction.
- 7.2.16. Add 20mL of 0.05M ammonium citrate to each Sr Resin cartridge reservoir to strip Pb. Engage vacuum. Allow solution to completely pass through each cartridge at 1mL/min. Set Pb samples aside for Pb source preparation.

- 7.3. Sample preparation for Po analysis: Auto Deposition on Ag, Cu or Ni planchet.
 - 7.3.1. Rinse polished planchets with water, ethanol and acetone. Let air dry.
 - 7.3.2. Place the planchets, polished side facing out, in the cap of a 50mL polypropylene centrifuge tube.

Note: The planchet should just fit inside the centrifuge tube cap and remain in place while fastening the cap to the centrifuge tube. If the planchet does not stay in place by friction alone, it can be affixed to the centrifuge tube cap using a hot glue gun.

- 7.3.3. Add 1mL of 1M ascorbic acid to each Po fraction. The ascorbic acid will reduce any Fe(III) in the solution and improve reduction of polonium and adhesion to the planchet.
- 7.3.4. Carefully place a cap, containing a planchet, on each sample tube. Seal the cap tightly and invert the sample tube so that the planchet is submerged below the solution containing polonium.

Note: Heating the samples in the following step can lead to some pressure buildup in the centrifuge tubes. A pressure release hole can be melted into the centrifuge tube with a soldering tool.

- 7.3.5. Place the samples in a stirring water bath or ultrasonic water bath. Heat to a temperature of 80-90°C and stir for at least 2 hours.
- 7.3.6. Remove tubes from water bath and carefully drain solution to waste.
- 7.3.7. Remove each planchet from the centrifuge tube cap and rinse it with water, ethanol and then acetone. Let planchets air dry.
- 7.3.8. Planchets may be heated on a 200-300°C hotplate for 15-30 minutes to convert Po to Po oxide to minimize the risk of detector contamination. However, care must be taken not to overheat planchets and lose polonium to volatilization.
- 7.3.9. Count in an alpha spectrometry system.
- 7.4. Sample preparation for Po analysis: Bismuth phosphate precipitation onto Resolve[®] filter.
 - 7.4.1. Add 125 μ L of bismuth carrier, 0.75mL of 3.2M (NH4)2HPO4, and 50 μ L 30% H₂O₂ to each polonium sample.

- 7.4.2. Mix well.
- 7.4.3. Add 0.6mL of concentrated NH4OH to each polonium sample.
- 7.4.4. Mix well.
- 7.4.5. Wait 20-30 minutes.
- 7.4.6. Collection bismuth phosphate precipitate, containing the polonium, on an Eichrom Resolve Filter as described in steps 5.4 to 5.15 of Eichrom Method SPA01.
- 7.5. Sample preparation and analysis option for Pb determination
 - 7.5.1. Planchet option
 - 7.5.1.1. Samples should be precipitated within an hour after eluting from resin as Bi may not fully co-precipitate with Pb.
 - 7.5.1.2. For each sample analyzed, clean a 2 inch diameter counting planchet by moistening a paper towel with ethanol, wiping the dish and letting it dry.

Note: Planchets can also be annealed in an oven at 450°C for 1.5 hours. Properly annealed planchets will appear bronze/brown in color. Do not overheat planchets or they could become more susceptible to acid degradation.

- 7.5.1.3. Weigh the dried counting planchet on an analytical balance and record the weight to 0.0001 gram.
- 7.5.1.4. Place each planchet in a fume hood, either under a heat lamp or on a hot plate with low heat.
- 7.5.1.5. Very carefully add 4mL of concentrated sulfuric acid to each Pb fraction.
- 7.5.1.6. Cap the tubes and mix the solution well. A white precipitate is formed. Mix periodically for 15-30 minutes to re-suspend the precipitate.
- 7.5.1.7. When the sample has cooled to room temperature, centrifuge and discard the supernate.
- 7.5.1.8. Add 10mL of water to the precipitate and mix well. Centrifuge the solution and discard the supernate. Repeat once.

- 7.5.1.9. Add 5mL of DI water to the precipitate and mix well to slurry.
- 7.5.1.10. Transfer the slurry to dried, weighed planchet in 2-3 mL aliquots. Evaporate to near dryness between additions.

Note: If the samples evaporate completely between additions, allow the planchets to cool slightly before adding more sample. This will minimize splattering and losses of the sample.

- 7.5.1.11. Rinse the empty sample tubes with 2-3 mL of water to recover any additional residue. Transfer the rinse to the planchet.
- 7.5.1.12. After all the solution has evaporated to dryness, cool each planchet.
- 7.5.1.13. Weigh each planchet, and record the weight to 0.0001 gram.
- 7.5.1.14. Cover planchet with aluminum foil and wait at least three days for ²¹⁰Bi ingrowth.
- 7.5.1.15. Count on a gas flow proportional counter.

Note: Samples should be counted with the aluminum foil covering the planchet. The aluminum foil will absorb the low energy beta emission from ²¹⁰Pb (60 keV). The same thickness of aluminum absorber (~0.03 inches) should be used for all samples and detector calibrations.

- 7.5.2. LSC/ICP-MS option.
 - 7.5.2.1. For LSC ²¹⁰Pb determination
 - 7.5.2.1.1. Add 7mL of the lead fraction from into a clean, labeled LSC vial.
 - 7.5.2.1.2. Add 15mL of appropriate scintillation cocktail to the vial. Shake to mix and ensure that cocktail does not from separate layers.
 - 7.5.2.1.3. Count on LSC after ²¹⁰Bi in-growth of at least 3 days. Take care to ensure that the measurement window is set to count the ²¹⁰Bi emission (~15-1200 keV) and not the entire beta spectrum.
 - 7.5.2.2. Analyze remaining Pb sample fraction and initial sample aliquot by AA/ICP/ICP-MS for Pb yield determination.



7.5.2.3. The Pb sample fraction can also be used to directly determine ²¹⁰Pb mass concentration via ICP-MS. (See reference 2).

8. CALCULATIONS

Polonium Tracer yield: ²⁰⁸Po/²⁰⁹Po

$$Po_{Y}$$
 (carrier yield) = $\frac{Po_{AF}}{Po_{AI} \times Po_{V}}$

where:

Po_{AF} = Activity of polonium tracer on final filter

 $Po_{AI} = Activity of polonium tracer spike solution$

 $Po_V = Amount of polonium tracer spiked into initial sample$

Calculate initial ²¹⁰Po activity

²¹⁰*Po* (pCi/L) =
$$\frac{S - B}{2.22 \times E \times Po_{Y} \times (e^{-\lambda(t_0 - t_{01})}) \times V}$$

where:

S = sample counts per minute B = background counts per minute E = efficiency of alpha counter Po_Y = Po tracer yield λ = 0.005008 day-1 (decay constant for ²¹⁰Po) t₀ = time of sample collection t₀₁ = time of mid-point of sample count V = volume of sample in liters

Gravimetric yield: Pb carrier - via planchet or filter

$$Pb_{Y}(carrier yield) = \frac{Rw - Tw - Bw}{Cw}$$

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- R_w = residue + planchet or filter, mg
- T_w = tare weight of planchet or filter, mg
- B_w = blank weight, mg
- C_w = PbSO4 added, mg

Chemical yield: Pb carrier - via ICP

$$Pb_{Y}(carrier yield) = \frac{C_F \times V_F}{C_I \times V_I}$$

where:

Cı	=	Pb initial concentration	
Vı	=	Initial Sample Volume at time of aliquot	
C _F =	Pb final concentration		
V _F =	Volume	e of Final Solution at time of final aliquot	

Calculate initial ²¹⁰Pb activity based on ²¹⁰Bi ingrowth

²¹⁰*Pb* (pCi/L) =
$$\frac{S - B}{2.22 \times E \times Pb_{Y} \times (1 - e^{-\lambda_{210Bi}(t_{L1} - t_{BS})}) \times V \times (e^{-\lambda_{210Pb}(t_{0} - t_{L1})})}$$

where:

S	=	sample counts per minute
В	=	background counts per minute
Е	=	efficiency of ²¹⁰ Bi emissions in beta counter
Pb_Y	=	Pb carrier yield
λ210В	i =	0.138 day-1 (decay constant for ²¹⁰ Bi)
t_{L1}	=	time of midpoint of sample count
t _{BS}	=	time of ²¹⁰ Bi separation, recorded in step 7.2.7
V	=	volume of sample in liters
λ210P	b=	0.0000855 day-1 (decay constant for ²¹⁰ Pb)
t ₀	=	time of sample collection

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9. References

- Horwitz, E. P., et al. "A lead-selective extraction chromatographic resin and its application to the isolation of lead from geological samples," *Analytica Chimica Acta*, 292, 263-273 (1994)
- 2) "Determination of Pb-210 in water using extraction chromatography," DOE Methods Compendium, RP280.
- 3) ASTM Method D7535-09, "Standard Test Method for Lead-210 in water."
- 4) Larivière, D., et al "Determination of 210Pb at ultra-trace levels in water by ICP-MS", *Analytica Chimica Acta*, 549(1-2), 188-196 (2005)
- 5) Maxwell, S.L., et al., "Rapid Determination of 210Po in Water Samples," *Radioanalytical and Nuclear Chemistry*, 298(3), 1977-1989, (2013)

(13) (14) (15) (16) Sr Resin Cartridge 2mL (17) (18) 14) Load sample on to Sr Resin. Allow liquid to drain to 17) Place clean labeled 50mL centrifuge tube below Sr Resin. Strip Po with 5mL 1M HNO₃. Allow solution Po sample to source preparation ← Add rinse to Sr Resin. Allow liquid to drain to waste. Pb sample to source preparation Waste to drain. Strip Po with 15mL 0.1M HNO₃. Allow 18) Place clean labeled 50mL centrifuge tube below Sr Resin. Strip Pb with 20mL 0.05M Precondition Sr Resin with 10mL 2M HCI. 15) Rinse centrifuge tube with 5mL 2M HCI. 16) Rinse Sr Resin with 5mL 2M HCl. Allow liquid to drain to waste. waste. Pb and Po are retained ammonium citrate. solution to drain. Ascorbic Sample HC N ⊒. Acid 7) After 30 minutes, turn off heat. Allow precipitate to settle or centrifuge 4) Cover with a watch glass and heat samples to near boiling for 1 hour. + 5) Turn down heat. Remove watch glass. Add 1mL of phenolphthalein Dissolve precipitate in 10mL 2M HCI. Add 1mL 1M ascorbic acid. Add Po yield tracer, 1mL of Pb carrier and 1mL of Fe carrier. 6) While stirring sample, slowly add conc. NH4OH until pH8-9. 10) Centrifuge ~10minutes at 2000rpm. Decant supernate. 11) Add 10mL DI water to precipitate. Mix well. Centrifuge. Centrifuge Transfer precipitate to centrifuge tube with DI water. 10 Aliquot up to 1000mL of water into glass beaker. 8) Decant supernate and discard as waste Iron Hydroxide ppt. 2) Acidify to pH 2 with conc. HNO₃ Water Sample in glass beaker. Acidify pH 2. Decant supernate. indicator.

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